



Effect of high hydrostatic pressure on the color and texture parameters of refrigerated Caiman (*Caiman crocodilus yacare*) tail meat

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ABSTRACT

The effect of applying high hydrostatic pressure (HHP) on the instrumental parameters of color and texture and sensory characteristics of alligator meat were evaluated. Samples of alligator tail meat were sliced, vacuum-packed, pressurized and distributed into four groups: control, treated with 200 MPa/10 min, 300 MPa/10 min and 400 MPa/10 min, then stored at $4\text{ }^{\circ}\text{C} \pm 1\text{ }^{\circ}\text{C}$ for 45 days. Instrumental color, texture profile and a sensory profiling using quantitative descriptive analysis were carried out on the 1st, 15th, 30th and 45th days of storage. HHP was shown to affect the color and texture of the product, and the sensory descriptors ($p < 0.05$). The results suggest that high pressure is a promising technology for the processing of alligator meat, especially low pressures (200 MPa) which can have positive effects on the quality of the product.

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1. Introduction

The breeding of caiman (*Caiman crocodilus yacare*) has been developing over the years in Brazil, thus representing an economically promising activity, especially in the Pantanal (swampland) region. Historically this activity has been related to manufacture of leather, but more recently the meat has been commercialized in specialized restaurants, with good acceptance (Vicente Neto et al., 2007).

As a consequence of market globalization, industry is searching for means to increase productivity and improve product quality, and thus new technologies are being developed and/or improved (Ferreira, Masson, & Rosenthal, 2008). High hydrostatic pressure (HHP) is a technology which is non-thermal, and consists of submitting the foods to pressures above 100 MPa (Cruz et al., 2010). This technology preserves the quality without significant alterations of the food matrix, with the advantage of efficiently eliminating microorganisms, providing microbiological safety and increased shelf life (Mathias et al., 2010). It has been used with success for meat products from different animal species (Aymerich, Picouet, & Monfort, 2008; Gou, Lee, & Ahn, 2010; Souza et al., 2011).

Among the adverse effects of high pressure, are alterations in color and texture, due to structural changes in macromolecules such as proteins. The covalent protein bonds are little affected by high pressure, but hydrophobic and electrostatic bonds can be affected, causing significant conformational changes and affecting functionality, frequently irreversibly, depending on the nature of the protein and the pressure applied (Lamballerie-Anton, Taylor, & Culioli, 2002). These attributes are important quality parameters and directly influence the consumer (Fletcher, Qiao, & Smith, 2000).

The application of preservation methods to alligator meat is a recent innovation (Vieira, 2010), but there are no available studies on the application of high hydrostatic pressure to this meat. Thus the objective of this study was to evaluate the effect of HHP on the quality parameters and sensory characteristics of alligator tail meat.

2. Material and methods

2.1. Sampling

Tail samples from 24 caiman (*Caiman crocodilus yacare*) were used, the caiman were reared in captivity to approximately 2.5 years of age, chosen at random and humanely slaughtered (Brasil, 2000) at the abattoir of the Swampland Alligator Breeder Cooperative (SIF 2452) in the city of Cáceres, State of Mato Grosso, Brazil. The 24 carcasses were cooled, the tails deboned and vacuum-packed (Criovac®), and

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transported to the Sensory Analysis Laboratory (UFF, Rio de Janeiro, Brazil) in isothermal boxes containing broken ice. They were then removed from the vacuum packs, cut into 25 g portions, packed into sterile plastic bags (10 × 4 cm), and transported in isothermal boxes containing ice to Embrapa Food Agro-industry – RJ, Brazil, where they were vacuum packed (gas-run, model 30 from Engevac©) and maintained at 4 °C ± 1 °C until pressurized.

2.2. Experimental design

Twenty-four alligator tails were vacuum-packed and divided into four groups each with six tails. Each group was divided at random into 25 g portions, giving a total of 20 samples per group, which were further divided into 4 sub-groups of 5 samples each (repetitions), submitted to the proposed treatments, and analyzed after four storage periods (1, 5, 30 and 45 days), giving a total of 80 sample units. The operational conditions for each treatment and the respective codes were control (CON), 200 (P200), 300 (P300) and 400 (P400) MPa pressure treatments for 10 min.

2.3. High hydrostatic pressure

The pilot equipment *Stansted Fluid Power* – model S-FL-850-9-W was used for pressurization. The pressure level was adjusted (200, 300 and 400 MPa) and the time of 10 min was controlled manually, maintaining the operational temperature at 20 °C. The come-up rate was approximately 300 MPa/min and total decompression took about 30 s. The samples were introduced into the stainless steel perforated cylinder of the equipment, whose dimensions were approx. 7.0 cm in diameter and 20.0 cm in length, using 70% alcohol as the pressure-transmission medium. After hermetically closing the chamber containing the cylinder, two pneumatic pumps were sequentially switched on to raise the pressure to the desired conditions. At the end of the cycle, the chamber was depressurized and opened to remove the samples.

2.4. Instrumental color analysis

The color parameters of L^* (luminosity), a^* ($-a$ = green; $+a$ = red) and b^* ($-b$ = blue; $+b$ = yellow) were obtained using a portable Konica Minolta model CR 400 colorimeter (Konica Minolta Sensing, Inc., Osaka, Japan). For the reading, the *in natura* samples were transversally cut (thickness of 1 cm) and maintained at room temperature for 30 min. The result was obtained from the mean of measurements made at three distinct regions of each sample.

2.5. Instrumental texture analysis

The texture analysis was carried out through texture profile analysis (TPA) (Bourne, 1978) under the following conditions: *in natura* samples cut into 1 cm³ cubes at a temperature of 10 °C, model TA-Hdi texturometer (Stable Micro System, London, England) with a 36 mm diameter cylindrical metal probe (P/36R), compression to 50% of the original height in two cycles, pre-test speed: 3.00 mm/s; test speed: 1 mm/s; and post-test speed: 3 mm/s, time between compressions: 2 s, and 100 g of force per area. The data were processed the Texture Expert for Windows (R, Stable Micro System), to give the cohesiveness, hardness, springiness and resistance. Ten repetitions were made for each group on each sampling day.

2.6. Sensory profiling

The sensory profile of each product was determined by eight selected and trained assessors, regular consumers of meat products, using the quantitative descriptive analysis method (QDA) developed by Stone, Sidel, Oliver, Woosley, and Singleton (1974). This analysis

was carried out using cooked and raw samples after 1, 15, 30 and 45 days storage at 4 °C. For the QDA, control and pressurized at 200, 300 and 400 MPa samples were cooked by immersion in water until the geometric center reached approximately 70 °C, which was monitored by digital thermometer. After cooking the samples were cut into 1 cm³ cubes and presented to the assessors.

The assessors were recruited with the aid of an individual oral interview amongst students already trained and used to this type of sensory analysis. The panel had already carried out this type of analysis and therefore had experience in the type of evaluation. Ten assessors took part in the test (five men and five women aged between 25 and 39), students of the postgraduate program in Veterinary Hygiene and Technological Processing of food of the Fluminense Federal University, previously orientated and trained in the analysis of control and pressurized samples.

During training of the sensory panel, the samples were offered to the assessors and the attributes of appearance, aroma, flavor and texture determined from an open discussion amongst the panel members, moderated by a leader. After determining the attributes, the panel met for a further six 2-hour sessions to establish, by consensus, the definitions and references for the subsequent elaboration of the scorecard. After identification of the attributes and definition of the references, training with the descriptive terms was carried out with anchor points of “slight” or “a lot” for each attribute evaluated. Before carrying out the QDA, the performance of the panel was evaluated, verifying their discrimination between samples, repeatability and agreement amongst the members (Damásio & Costell, 1991). Analysis of variance (ANOVA) was used for this purpose, with two causes of variation (sample and repetition) for each attribute and assessors, selecting those assessors with significant F sample values ($p < 0.30$) and non-significant F repetition values ($p > 0.05$). The eight assessors selected (three men and five women) took part in the subsequent tests. Four repetitions were used per treatment (control and pressurized sample) of the alligator tail meat and the samples were served in a monadic way, coded with three-algorithm numbers and with a balanced presentation order.

For the final evaluation of all the attributes the samples were presented at room temperature on disposable white plastic plates under white light in individual booths. Salted bread and mineral water at room temperature were offered to clean the palate between samples. The trained panel carried out the QDA of the samples under laboratory conditions with five repetitions per assessor, using a scorecard with a non-structured 15 cm-long perception intensity scale.

2.7. Statistical analysis

A 4 × 4 factorial analysis of variance was carried out according to the pressure level (control, 200, 300 and 400 MPa), days of storage (1, 15, 30 and 45) and the interaction between the variables. Results showing a significant effect of pressure and/or storage were tested by ANOVA according to a completely random design for time and pressure level separately, followed by Tukey test ($p < 0.05$). The software “Statistical Analysis System” (SAS, 2000) was used to carry out these analyses.

The results of the quantitative descriptive analysis were evaluated by principal components analysis in a correlation matrix with the data centered on the mean. A matrix was elaborated with 4 lines and 5 columns, the lines representing the samples, and the columns the sensory descriptors. Hierarchical Clustering Analysis (HCA, Souza et al., 2011) was also carried out with the objective of evaluating the separation of the samples with respect to the QDA attributes. The clustering parameters were: dissimilarity, Euclidean distance, agglomeration method, Ward's method and manual truncation. Finally the parameters involved in the instrumental analyses of color and texture profile were related through Pearson's correlation. All these analyses were carried out using the software XLSTAT for Windows 2010.

3. Results and discussion

3.1. Instrumental color

Table 1 shows the values obtained for instrumental color for the samples submitted to high hydrostatic pressure. In general high hydrostatic pressure had an effect on the parameters ($p < 0.05$), the effect being directly proportional to the values for L^* (69.75–79.54) and inversely proportional to the values for a^* (5.15–8.48) when compared to the control group, which had variations of 65.43 to 67.47 and 7.96 to 10.30 for these two parameters, respectively.

The loss of color caused by the process can be explained by the oxidation of myoglobin, with a consequent decline in a^* (Carlez, Veciana-Nogues, & Cheftel, 1995), while the alterations in L^* can be explained by changes in the myofibrillar and sarcoplasmatic proteins, resulting in alterations to the surface of the meat (Jung, Ghoul, & Lamballerie-Anton, 2003). The results were similar to those observed in other food matrixes such as turbot fillets (Chevalier, Bail, & Ghoul, 2001), oysters (Cruz-Romero, Smiddy, Hill, Kerry, & Kelly, 2004) and fresh beef (Picouet, Pérez-Juan, & Realini, 2008).

During storage the L^* values of all treatments showed a slight increase by the 45th day, although P200 was the only treatment that significantly increased the value ($p < 0.05$). a^* for the control group increased, while P300 and P400 it decreased by day 45 ($p < 0.05$). b^* values did not change significantly ($p > 0.05$) between the first and the last day of storage in all treatments. According to Jung et al. (2003), the changes in color of the meat during storage could be associated with both enzymatic and non-enzymatic reactions, resulting in degradation of myofibrillar proteins and disorganization of the myofibrils. Similar results were observed with carp fillets (Sequeira-Munoz, Chevalier, Lebail, Romaswamy, & Simpson, 2006) and sea bass fillets (Chéret, Chapleau, Delbarre, & Verrez-Bagnis, 2005), submitted to pressures similar to those used in the present study.

3.2. Texture profile

Table 2 shows the results obtained in the texture profile. Changes were observed in the values for cohesiveness, springiness and resistance, which gradually increased with increased pressure applied ($p < 0.05$). With respect to storage time, random behavior was observed for all pressures except 400 MPa which showed constant values ($p > 0.05$). For hardness, the smallest values were observed for the samples pressurized at 200 MPa over the whole storage period.

Ma and Ledward (2004) reported that structural alterations in the contractile myofibrillar proteins are the main factor responsible for

Table 2

Mean and standard deviations of texture parameters cohesiveness, hardness, springiness and resistance of treated caiman tail meat during refrigerated storage (1, 15, 30 and 45 days).

Days of storage	CON	P200	P300	P400
Cohesiveness				
1	0.41 ^{cAB} ± 0.05	0.46 ^{bAB} ± 0.02	0.50 ^{aA} ± 0.02	0.49 ^{abA} ± 0.05
15	0.40 ^{bB} ± 0.05	0.47 ^{aA} ± 0.03	0.49 ^{aA} ± 0.04	0.51 ^{aA} ± 0.04
30	0.35 ^{cC} ± 0.03	0.44 ^{bB} ± 0.03	0.48 ^{aA} ± 0.04	0.51 ^{aA} ± 0.02
45	0.42 ^{bA} ± 0.03	0.45 ^{abAB} ± 0.03	0.47 ^{aA} ± 0.06	0.48 ^{aA} ± 0.05
Hardness (N)				
1	17.41 ^{aA} ± 1.70	12.63 ^{bA} ± 1.55	15.88 ^{aA} ± 2.43	16.20 ^{aA} ± 1.94
15	14.47 ^{abB} ± 2.13	12.22 ^{bA} ± 2.05	14.55 ^{abAB} ± 1.70	14.96 ^{abB} ± 2.23
30	12.88 ^{abC} ± 2.02	8.90 ^{bb} ± 2.23	12.10 ^{abC} ± 1.82	13.02 ^{ab} ± 2.32
45	11.91 ^{aC} ± 2.12	7.98 ^{bb} ± 1.77	11.93 ^{aC} ± 2.51	12.77 ^{ab} ± 2.52
Springiness				
1	0.52 ^{cA} ± 0.05	0.64 ^{bA} ± 0.04	0.79 ^{aA} ± 0.05	0.76 ^{aA} ± 0.03
15	0.43 ^{cBC} ± 0.05	0.62 ^{bA} ± 0.05	0.77 ^{abB} ± 0.03	0.77 ^{aA} ± 0.05
30	0.36 ^{dC} ± 0.04	0.48 ^{cb} ± 0.05	0.66 ^{bC} ± 0.06	0.77 ^{aA} ± 0.05
45	0.49 ^{cAB} ± 0.04	0.58 ^{bA} ± 0.04	0.76 ^{abB} ± 0.05	0.72 ^{aA} ± 0.05
Resistance				
1	0.24 ^{bA} ± 0.03	0.26 ^{abA} ± 0.03	0.25 ^{abA} ± 0.04	0.28 ^{aA} ± 0.03
15	0.17 ^{bb} ± 0.05	0.19 ^{bb} ± 0.05	0.21 ^{ba} ± 0.02	0.27 ^{aA} ± 0.05
30	0.16 ^{cb} ± 0.03	0.24 ^{baB} ± 0.04	0.22 ^{ba} ± 0.04	0.31 ^{aA} ± 0.05
45	0.19 ^{bb} ± 0.03	0.23 ^{abAB} ± 0.05	0.23 ^{abA} ± 0.05	0.29 ^{aA} ± 0.07

a, b, c, A, B, C Different letters within column and lines indicate significant differences among treatments ($p < 0.05$). a, b, c represent pressures in lines and A, B, C represent storage period, in columns.

texture changes. In the pressure range from 100 to 300 MPa, the changes are normally reversible, whereas at higher pressures they are normally non-reversible (Rastogi, Raghavarao, Balasubramaniam, Niranjana, & Knorr, 2007). The lysosomes rupture at pressures around 200 MPa, promoting an increase in autolytic activity and tenderization of the meat (Lamballerie-Anton, Taylor, & Culioli, 2002).

Chéret et al. (2005) observed a decrease in hardness for samples pressurized at 200 MPa, with a subsequent increase proportional to the increase in pressure, as also reported by Ashie and Simpson (1996) for bluefish and sheephead fish samples. Angsupanich and Ledward (1998) reported that the muscle of cod (*Gadus morhua*) showed an increase in springiness when submitted to pressures of 400 and 600 MPa for 20 min. Yagiz, Kristinsson, Balaban, and Marshall (2007), studying the effect of high pressures on rainbow trout, found that cohesiveness was significantly higher in samples submitted to pressures between 300 and 600 MPa.

3.3. Sensory profiling

Tables 3 and 4 show the mean results obtained for the sensory descriptors in the quantitative descriptive analysis, and also the description for each attribute. Storage time did not affect the sensory attributes (data not shown), thus the differences between treatments were caused by the pressure level applied. Six sensory descriptors were established and evaluated by the sensory panel, as follows: the raw meat color, color of the cooked meat (appearance), alligator meat flavor (flavor), tenderness, succulence, fibrosity and cohesiveness (texture). There was no consensus amongst the sensory panel with respect to any sensory descriptor related to aroma, and thus it was excluded from the sensory profile.

Of the sensory descriptors, only the attribute “raw color” showed relevant differences ($p < 0.05$), suggesting that the use of HHP caused a loss of color, explained by oxidation of the myoglobin, with a consequent decline in the values for a^* (Carlez et al., 1995). The attribute “cooked color” distinguished all treatments from samples submitted to 400 MPa, which had the lowest score ($p < 0.05$). For the attribute “tenderness”, the highest mean was obtained with the application of 200 MPa, although there was no statistically significant difference when compared with the other treatments ($p > 0.05$). No effect on

Table 1

Means and standard deviations of L^* , a^* and b^* from control pressurized caiman raw tail meat during refrigerated storage (1, 15, 30 and 45 days).

Days of storage	CON	P200	P300	P400
L^* raw				
1	65.43 ^{cA} ± 1.66	69.75 ^{bb} ± 1.23	78.76 ^{aA} ± 1.21	78.44 ^{aA} ± 0.75
15	66.09 ^{cA} ± 2.95	70.12 ^{bb} ± 0.81	77.19 ^{aA} ± 1.50	78.76 ^{aA} ± 2.52
30	65.59 ^{cA} ± 1.25	71.07 ^{bb} ± 1.81	78.05 ^{aA} ± 2.98	77.71 ^{aA} ± 1.99
45	67.47 ^{cA} ± 1.82	74.86 ^{bA} ± 1.15	79.29 ^{aA} ± 2.55	79.54 ^{aA} ± 2.17
a^* raw				
1	8.53 ^{abc} ± 0.17	7.87 ^{abAB} ± 0.64	7.13 ^{bca} ± 0.60	6.80 ^{cA} ± 0.38
15	7.96 ^{bc} ± 0.40	7.24 ^{abB} ± 0.47	6.41 ^{bAB} ± 0.13	5.61 ^{cb} ± 0.37
30	8.47 ^{ab} ± 0.28	8.48 ^{aA} ± 0.36	5.75 ^{bBC} ± 0.71	5.73 ^{bb} ± 0.28
45	10.30 ^{aA} ± 0.67	7.22 ^{bb} ± 0.30	5.40 ^{cc} ± 0.31	5.15 ^{cb} ± 0.63
b^* raw				
1	8.76 ^{bA} ± 0.54	9.36 ^{abA} ± 0.25	9.15 ^{ba} ± 0.42	10.02 ^{aA} ± 0.61
15	9.10 ^{aA} ± 0.38	8.70 ^{aA} ± 0.48	8.94 ^{aA} ± 0.67	9.28 ^{abB} ± 0.27
30	7.85 ^{bb} ± 0.50	9.28 ^{aA} ± 0.57	9.06 ^{aA} ± 0.40	9.01 ^{ab} ± 0.54
45	8.40 ^{bAB} ± 0.42	8.96 ^{abA} ± 0.57	9.33 ^{aA} ± 0.37	9.25 ^{abB} ± 0.14

a, b, c, A, B, C Different letters within column and lines indicate significant differences among values ($p < 0.05$). a, b, c represent pressures, in other words, lines; and A, B, C stand for storage period, in columns.

Table 3
Means and standard deviations of sensory descriptors (raw color, cooked color, flavor, tenderness, succulence, fibrosity and cohesiveness) of caiman tail meat following pressure treatments.

Treat.	Raw color	Cooked color	Flavor	Tenderness	Succulence	Fibrosity	Cohesiveness
CON	7.72 ^a ± 1.31	2.27 ^a ± 0.91	12.60 ^a ± 1.70	9.51 ^a ± 1.51	10.31 ^a ± 1.19	5.97 ^a ± 2.10	6.00 ^a ± 2.23
P200	5.14 ^b ± 1.28	1.37 ^a ± 0.71	12.26 ^a ± 1.71	10.71 ^a ± 2.89	8.70 ^a ± 1.55	5.25 ^a ± 2.48	5.34 ^a ± 0.75
P300	2.12 ^c ± 1.01	1.39 ^a ± 0.98	12.59 ^a ± 2.65	9.35 ^a ± 2.39	9.83 ^a ± 1.52	6.05 ^a ± 2.07	6.06 ^a ± 1.83
P400	1.41 ^c ± 1.01	0.96 ^b ± 0.26	12.42 ^a ± 1.82	9.83 ^a ± 2.94	9.25 ^a ± 2.31	5.73 ^a ± 1.74	6.20 ^a ± 2.43

a, b, c Different letters within column and lines indicate significant differences among treatments ($p < 0.05$).

Table 4
Sensory attributes and reference standards used to anchor the panel scores.

Attribute	Definition	References
Raw meat color	Pinkish white	Light = cooked fish (<i>Rhinobatidae</i> family) fillet Dark = raw chicken breast
Cooked meat color	Very light gray	Light = cooked fish (<i>Rhinobatidae</i> family) fillet Dark = cooked pork loin (<i>M. Longissimus dorsi</i>)
Alligator meat flavor	<i>Sui generis</i>	Slight = cooked fish (<i>Rhinobatidae</i> family) fillet A lot = cooked alligator tail meat
Tenderness	Strength needed to cut the sample at the first bite	Slight = cooked beef (<i>M. Semitendinosus</i>) A lot = cooked fish (<i>Rhinobatidae</i> family) fillet
Succulence	Amount of juice expelled during chew	Slight = cooked fish (<i>M. Semitendinosus</i>) A lot = cooked fish (<i>Rhinobatidae</i> family) fillet
Fibrosity	Shape and fiber orientation during chew (geometric propriety)	Slight = cooked fish (<i>Rhinobatidae</i> family) fillet A lot = cooked beef (<i>M. Semitendinosus</i>)
Cohesiveness	Rate which sample particles remain together	Slight = cooked fish (<i>Rhinobatidae</i> family) fillet A lot = cooked beef (<i>M. Semitendinosus</i>)

flavor was observed ($p > 0.05$), which could be related to the minimal production of low molecular weight compounds, especially those responsible for the flavor of alligator meat, as reported by Telléz-Luis,

Ramírez, Pérez-Lamela, Vasquéz, and Simal Gándara (2001). Succulence was more pronounced in the control sample, whereas those treated at 400 MPa had greater cohesiveness, and those treated with 300 MPa, greater fibrosity, although these differences were not statistically significant ($p > 0.05$). HHP processing affects the structure of the protein molecules, promoting water expulsion and protein aggregation that reduces succulence and increases cohesiveness and fibrousness (Dong Sun & Holley, 2010). Although this has also been observed in other meats (Crehana, Troya, & Buckley, 2000; Suzuki, Kim, Tanji, Nishiumi, & Ikeuchi, 2006; Zbigniew et al., 2011), the present study does not support these findings.

The principal components analysis (PCA, Fig. 1) and the hierarchal clustering analysis (HCA, Fig. 2) confirmed the results discussed above. A total of 96.49% of the variability in the data was explained by the PCA, 64.24% of the variation being demonstrated in the first principal component (PC1) and 32.26% in the second (PC2). Although the first principal component showed the greatest percentage of explanation (64.24%), it can be seen that only in the second principal component were the samples clearly separated. The formation of groups can be observed: the control samples and samples submitted to 200 MPa (CON and P200) were in the first group, and the samples submitted to 300 MPa and 400 MPa (P300 and P400) were in the second group. It can be seen that the sensory descriptors of tenderness, raw meat color, cooked meat color and succulence were responsible for the differentiation between the samples, notably the tenderness for samples submitted to lower pressures. In the case of the HCA dendrogram (Fig. 2), the existence of two segments was clear: one containing the samples CON and P200 and the other containing the P300 and P400 samples.

Results from the instrumental analyses of color and texture and those from the sensory analyses were negatively correlated ($r = -0.91$) as can

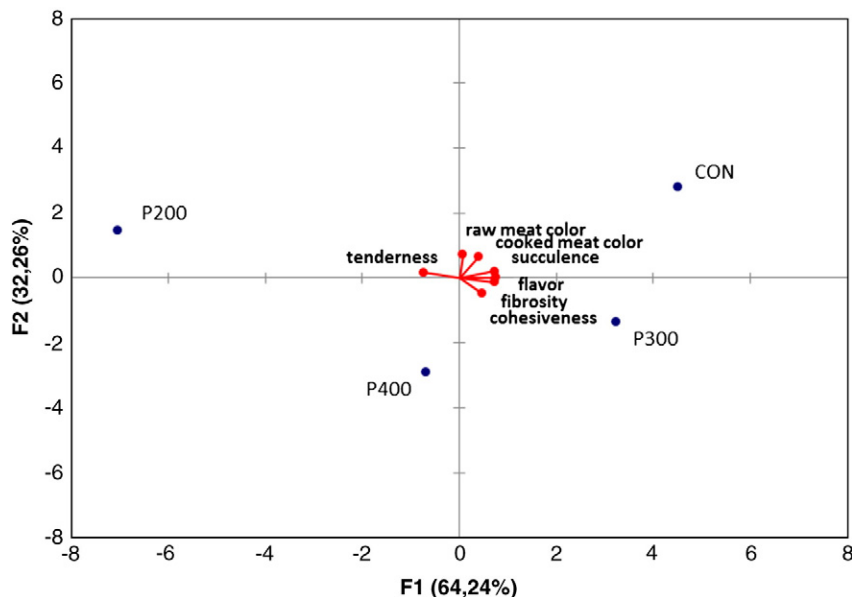


Fig. 1. Principal component analysis of sensory descriptors according to the QDA panel of pressurized alligator meat.

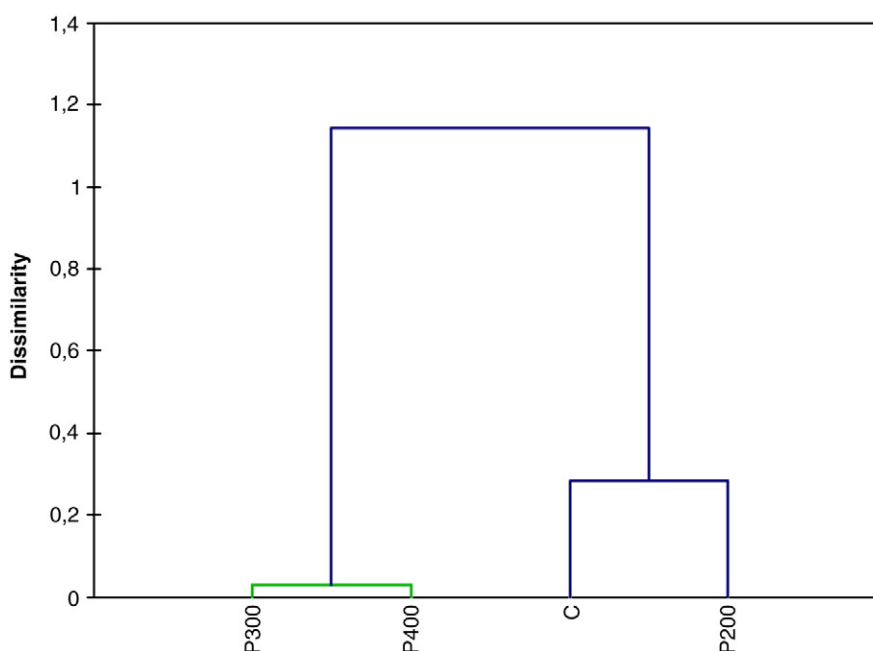


Fig. 2. Dendrogram of the hierarchical cluster analysis (HCA) of samples assessed by the QDA panel.

be seen between the instrumental analysis for hardness and the sensory analysis for tenderness, a similar result was obtained by Freitas (2005). The instrumental and sensory parameters of cohesiveness showed a positive correlation of 0.90. In the descriptive sensory analysis, the color intensity scale varied in the opposite direction from the raw L^* and b^* scales, for which high scores signified less luminosity and less yellowness. Thus the values for the instrumental color analysis (values of L^* and b^*) and the sensory analysis were negatively correlated, ($r = -0.99$ and -0.91), respectively. On the other hand, a^* for raw meat, redness, was positively correlated ($r = 0.99$) between the instrumental and sensory data, indicating it is a parameter with great influence on the differentiation of the raw color.

In general it can be seen that even when using lower pressures, there was a positive alteration in the sensory characteristics of the alligator meat, especially with respect to color, flavor and tenderness, which was also shown to be relevant in other sensory studies involving meat products submitted to high hydrostatic pressure (Sorenson et al., 2011). Future studies should include tests with consumers, and also the effect of receiving information concerning the high pressure process, on sample acceptance.

4. Conclusions

The use of high hydrostatic pressure in the processing of alligator meat showed promising results. There was good correlation between instrumental and sensory analysis of color and texture. Pressurization at 200 MPa was considered the best and most viable treatment under the conditions of this research demonstrating the lowest modification of lightness and redness and decreased hardness. Future studies should emphasize consumer tests and also other physicochemical parameters of importance for product quality, such as lipid oxidation. In addition food safety studies should be carried out with respect to the inactivation capacity and survival of pathogenic microorganisms.

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